# The use of Hyaluronic Acid as an adjuvant Therapeutic Approach to Non-**Surgical Periodontal Therapy for Periodontitis**

# O uso do Ácido Hialurônico como uma abordagem terapêutica adjuvante à Terapia Periodontal Não Cirúrgica da Periodontite

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### **ABSTRACT**

Periodontitis is a highly prevalent infectious-inflammatory disease, characterized by alveolar bone resorption. Hyaluronic acid (HA) has stood out for its anti-inflammatory and antioxidant effects. The present study evaluated, through a literature review, the use of HA as an adjunctive therapeutic approach to non-surgical periodontal therapy (NSPT) of periodontitis (PD), carried out through scaling and root planing. For this, the Pubmed and Medline databases were used with the following descriptors: Hyaluronic Acid AND Periodontitis. Among 58 manuscripts found, 11 studies were selected after analyzing their title and respective abstracts. Pre-clinical or clinical research published from 2010 to 2020, including the use of HA exclusively in the NSPT of PD, were included for this literature review. Studies using other pharmacological interventions in conjunction with HA, other literature reviews and research related to the treatment of gingivitis, periodontal or pulp regeneration, surgical periodontal therapy, apical or peri-implantitis periodontitis were excluded. The main data of the selected manuscripts were

compiled in a table and, then, their results were compared and discussed among themselves. Despite a wide variety of methodological protocols carried out by prospective researches, it is noted that HA has potential healing, anti-inflammatory and antimicrobial effects, which can be interesting from the point of view of periodontal treatment, considering the infectious and inflammatory character of PD. There are also different commercial formulations of HA on the market. It is suggested, therefore, that HA can be promising as a chemical adjuvant to PD NSPT, and can be used in the form of subgingival irrigation in deep periodontal pockets.

**Descriptors:** Hyaluronic acid; Periodontitis.

### **RESUMO**

A periodontite é uma doença infectoinflamatória de alta prevalência, caracterizada pela reabsorção óssea alveolar. O ácido hialurônico tem se destacado por seus efeitos antoinflamatório e antioxidante. O presente estudo avaliou, por meio de uma revisão de literatura, o uso do HA como uma abordagem terapêutica adjuvante à terapia periodontal não cirúrgica (NSPT) da periodontite (PD), realizada através de raspagens e alisamento radiculares. Para isso, utilizaram-se as bases de dados Pubmed e Medline com os seguintes descritores: Hyaluronic Acid AND Periodontitis. Dentre 58 manuscritos encontrados, foram selecionados 11 estudos após análise de título e seus respectivos resumos. Foram incluídos, para a presente revisão de literatura, pesquisas pré-clínicas ou clínicas publicadas do período de 2010 a 2020, relacionando o uso exclusivamente do HA no NSPT da PD. Foram excluídos estudos utilizando outras intervenções farmacológicas conjuntamente ao HA, outras revisões de literaturas e pesquisas relacionadas ao tratamento da gengivite, à regeneração periodontal ou pulpar, à terapia periodontal cirúrgica, à periodontite apical ou peri-implantite. Os dados principais dos manuscritos selecionados foram compilados em tabela e, em seguida, os seus resultados foram confrontados e discutidos entre si. Apesar de uma ampla variedade nos protocolos metodológicos realizados pelas pesquisas prospectivas, nota-se que o HA apresenta potenciais efeitos cicatrizante, anti-inflamatório e antimicrobiano, os quais podem ser interessantes do ponto de vista do tratamento periodontal, considerando o caráter infectoinflamatório da PD. Existem, ainda, no mercado, diferentes formulações comerciais do HA. Sugere-se, portanto, que o HA pode ser promissor como adjuvante químico à NSPT da PD, podendo ser utilizado sob a forma de irrigação subgengival em bolsas periodontais profundas.

**Descritores:** Ácido hialurônico; Periodontite.

### 1 INTRODUCTION

Periodontal diseases are often pathologies of an infectious and inflammatory character, which affect individuals of varying ages and worldwide. Namely, in view of the accumulated presence of dental biofilm, composed of a varied and organized microbiota, an immune-inflammatory response occurs in the host with primary defense function. However, it has been well clarified in the literature that its exacerbation, characterized by leukocyte activation and massive release of inflammatory mediators, such as tumor necrosis factor (TNF)  $-\alpha$ , interleukin

(IL) -1 $\beta$ , IL-6, in addition to prostaglandin (PG) -E2, has been responsible for most of the tissue destruction observed<sup>1,2</sup>.

Thus, periodontitis (PD), which occurs after the apical extension of the inflammatory process before restricted protection periodontium, presents clinical signs and irreversible sequelae associated with alveolar bone resorption (ABR) and the destruction of collagen, such as the formation of periodontal pockets, increased probing depth, bleeding on probing and apical migration of the junctional epithelium<sup>1,2,3</sup>.

Interestingly, the ABR, a characteristic of PD, is similar to the bone resorption observed in some chronic systemic inflammatory diseases, such as osteoarthritis, in which focal bone erosion occurs in the affected joints<sup>4</sup>. In fact, it has been described that there is a close relationship between the immune and bone systems, in such a way that chronic inflammatory processes predispose to bone destruction, which explains the intensified ABR in PD as the immunoinflammatory process spreads in the constant presence of periodonto-pathogens. Thus, this being considered the initiating factor of all periodontal destruction, non-surgical periodontal therapy (NSPT) of PD involves mechanical debridement of biofilm and dental calculus by means of scaling and root planing (SRP) and patient training in oral hygiene that control or reduce the presence of biofilm<sup>5,6</sup>.

Although SRP allows numerous benefits, such as a reduction in the concentration of microorganisms and their products, a decrease in the depth of bags, reduction of inflammation and edema with rehabilitation of the junctional epithelium, it also has certain limitations. These can be anatomical, related to the professional or related to the patient, such as irregularities in the dental surface, difficulty in accessing very deep pockets, bifurcation areas, among others. Thus, chemical adjuvants, especially those with immunomodulatory and antimicrobial capabilities, when associated with PD NSPT, may be relevant in enhancing the patient's responsiveness through such therapy, reducing periodontal sequelae and the evolution of ABR<sup>7</sup>.

Among the pharmacological interventions studied for a few decades for this purpose, hyaluronic acid (HA) stands out, a high molecular weight glycosaminoglycan and a fundamental component of extracellular matrices that contributes to tissue hydrodynamics, and is naturally involved in migration processes and cell proliferation<sup>8,9</sup>.

HA has demonstrated anti-inflammatory effect in several areas of medicine<sup>10</sup>. In the treatment of burns, it has been used in order to enhance the healing process<sup>11</sup>. In fact, the protective effect of AH against joint degeneration and synovial inflammation has been described in an experimental model of osteoarthritis<sup>12</sup>. In fact, its promising activity in the treatment of

osteoarthritis has been observed constantly<sup>13,14</sup>. As for its antimicrobial effect, it has been demonstrated against specimens of *Escherichia Coli* (*E. coli*) and *Staphylococcus Aureus* (*S. aureus*)<sup>15</sup>. In addition, the ability to reduce the formation of bacterial biofilms has also been investigated in the literature<sup>16</sup>.

Thus, considering that PD is characterized by infectious etiology and inflammatory ABR, coupled with the knowledge that HA has stood out for its biological potentials, especially anti-inflammatory, antimicrobial and bone protector, the present literature review is important for to evaluate results of scientific research related to the adjuvant potential of this drug in NSPT of PD.

### 2 MATERIALS AND METHODS

For the present literature review, scientific articles published from 2010 to 2020 were searched in the Pubmed and MEDLINE databases, with the following descriptors, according to the Health Sciences Descriptors (DeCS): Hyaluronic Acid AND Periodontitis.

Among 58 manuscripts found, 11 scientific articles were selected. For the selection of these, abstracts of those published from the year 2010 were read and only articles from preclinical or clinical scientific research were selected, relating the use of exclusively HA in the NSPT of the PD. Thus, studies using other pharmacological interventions in conjunction with AH, other literature reviews and research related to the treatment of gingivitis, periodontal or pulp regeneration, non-surgical periodontal therapy, apical periodontitis or peri-implantitis were excluded. The 11 selected scientific articles were read in full and their data were compiled in table and then compared with each other based on the scientific literature.

### **3 RESULTS**

As can be seen in table 01, the results of the 11 studies selected for the present literature review are compiled (Table 01). It was observed that 10 studies were prospective clinical trials conducted in patients with periodontitis<sup>17-27</sup>, in which the administration of HA showed additional benefits to NSPT, significantly reducing, compared to sites or control groups, Probing Pocket Depht (PPD)<sup>17,18,20-23, 25</sup>, Clinical Attachment Level (CAL)<sup>20-23</sup>, Gingival Index (GI)<sup>17,19,21</sup>, Plaque Index (PI)<sup>20,21</sup> and Bleeding on Probing (BoP)<sup>17-20,24</sup>.

Of these, the majority<sup>17-24</sup> selected patients with periodontitis, whose periodontal analyzes were performed and, in each, intraoral sites were chosen: control and test, which received,

respectively, NSPT and placebo or NSPT and subgingival application of HA in different concentrations.

In another 2 clinical trials<sup>25,26</sup>, it was observed that there was a random division of patients into control and test groups, which could increase the bias of some parameters analyzed, especially those that were closely related to the patient's hygiene capacity, considering that manual skill and oral hygiene habits can vary considerably between individuals. In fact, in these 2 studies, few significant differences regarding the clinical periodontal parameters evaluated were found for the test group, compared to the control group.

Other analyzes consisted of biochemical evaluations, performed through the collection of crevicular fluid, the levels of bacterial specimens, Myeloperoxidase (MPO), Calprotectin, Neutrophil elastase (NE) and  $\beta$ -defensin-2 (hBD-2). As for antimicrobial effects, it was observed that the administration of HA significantly reduced the concentration of bacterial specimens in the crevicular fluid<sup>20,25</sup>. However, it was observed that the HA did not alter, compared to the control sites, the levels of MPO<sup>18,25</sup>, Calprotectin<sup>18</sup> and NE<sup>21</sup>, however significantly increased the levels of hBD-2<sup>23</sup>.

In an in vitro study, the results were also promising<sup>27</sup>. In general, the beneficial results found resulted from the healing, anti-inflammatory and antimicrobial effects of HA. No preclinical research on animals was found.

Table 1 - Studies selected for the construction of the article.

**Methodology:** At baseline, patients with periodontitis were evaluated for periodontal parameters and, in each, two intraoral regions were chosen: control and test, which received, respectively, NSPT and placebo or NSPT and subgingival application of HA. Then, patients were reassessed at different times.

| Authors    | Reviews                                 | Results                         |
|------------|---|---------------------------------|
| Pilloni et | Periodontal parameters evaluated:       | During the 3 weeks, HA          |
| al., 2011  | Probing Pocket Depht (PPD), Gingival    | significantly and progressively |
|            | Index (GI), Bleeding on Probing         | reduced the periodontal         |
|            | (BoP), plaque index (PI) and Clinical   | parameters evaluated, with the  |
|            | Attachment Level (CAL).                 | exception of PI and CAL, in     |
|            | HA (HYAFF®, Anika Therapeutics          | which no significant            |
|            | Sr) was applied subgingivally daily for | differences were observed       |
|            | 3 consecutive weeks immediately after   | between the control and test    |
|            | non-surgical periodontal therapy        | sites.                          |
|            | (NSPT).                                 |                                 |
|            | Patients were assessed at baseline and  |                                 |
|            | 1, 2 and 3 weeks after of NSPT.         |                                 |

| Bevilacqua   | Periodontal parameters evaluated:      | At 45 days, HA significantly     |
|--------------|--|----------------------------------|
| et al., 2012 | PPD, BoP, PI and CAL.                  | reduced PPD and BoP at sites     |
|              | Parameters evaluated in gingival       | that received HA, compared to    |
|              | crevicular fluid (GCF):                | control. Although the other      |
|              | Myeloperoxidase (MPO) and              | evaluated periodontal            |
|              | Calprotectin levels.                   | parameters also showed           |
|              | HA (Aminogam Ò A, lotto 190308A,       | reductions, these were not       |
|              | Errekappa Euroterapici Spa, MI, Italy) | significant. As for the volume   |
|              | was applied subgingivally only once    | of crevicular fluid, significant |
|              | after the NSPT.                        | differences were found           |
|              | Patients were assessed at baseline and | between the sites after 7 and 45 |
|              | 7, 45 and 90 days after NSPT.          | days of reevaluation. The        |
|              |  | levels of MPO and                |
|              |  | Calprotectin did not show        |
|              |  | significant differences between  |
|              |  | the control and test sites.      |
| Gontiya e    | Periodontal parameters evaluated:      | HA significantly reduced IG      |
| Galgali,     | PPD, BoP, PI and CAL.                  | and BoP only after the 6th       |
| 2012         | HA (Gengigel®; 0.2% formulation)       | week after the start of          |
|              | was applied subgingivally once a week  | treatment. No additional         |
|              | immediately after NSPT, for 3 weeks.   | advantages were observed for     |
|              | Patients were assessed at baseline and | the administration of HA         |
|              | 4, 6 and 12 weeks after NSPT.          | regarding PPD and CAL in the     |
|              |  | three reevaluations.             |
|              |  | Histologically, there were also  |
|              |  | no significant differences       |
|              |  | regarding the inflammatory       |
| 5 1 11       |  | infiltrate.                      |
| Polepalle et | Periodontal parameters evaluated:      | The HA provided a significant    |
| al., 2015    | PPD, GI, BoP and CAL.                  | improvement in the periodontal   |
|              | Subgingival plaque samples were        | clinical parameters analyzed,    |
|              | collected immediately and 2 weeks      | with the exception of the        |
|              | after the start of NSPT for culture    | reevaluation period in the first |
|              | (CFU).                                 | week. In addition, there was a   |
|              | HA (0.8% formulation) was applied      | possible antimicrobial activity  |
|              | subgingivally once just after the      | of HA.                           |
|              | NSPT.                                  |                                  |
|              | Patients were assessed at baseline and |                                  |
| Mo11:1 '     | 1, 4 and 12 weeks after NSPT.          | The record AII and I also        |
| Mallikarjun  | Periodontal parameters evaluated:      | The use of AH reduced the        |
| et al., 2016 | PPD, GI, PI and CAL.                   | clinical periodontal parameters, |
|              |  | but without significant          |

|   | Parameters evaluated in the GCF:             | differences regarding the levels                       |  |  |
|---|--|--|--|--|
|   | Levels of neutrophil elastase (NE).          | of NE. This result shows that                          |  |  |
|   | HA (Gengigel®; 0.2% formulation)             | there is no positive correlation                       |  |  |
|   | was applied subgingivally once just          | between improvements in the                            |  |  |
|   | after NSPT.                                  | periodontal clinical parameters                        |  |  |
|   | Patients were assessed at baseline and       | evaluated after 6 weeks and the                        |  |  |
|   | 6 weeks after NSPT.                          | levels of NE.  |  |  |
| Shah et al.,  | Periodontal parameters evaluated:            | HA significantly improved the                          |  |  |
| 2016  | PPD, GI, PI and CAL.                         | periodontal parameters PPD                             |  |  |
|   | HA (Gengigel®; 0.8% formulation)             | and CAL, compared to the                               |  |  |
|   | was applied subgingivally once just          | control sites. No significant                          |  |  |
|   | after the NSPT.                              | differences were observed for                          |  |  |
|   | Patients were assessed at baseline and       | the other parameters evaluated.                        |  |  |
|   | 4 and 12 weeks after NSPT.                   | -  |  |  |
| Al-   | Periodontal parameters evaluated: PI,        | After 12 weeks, HA reduced                             |  |  |
| shammari,   | papillary bleeding index, PPD and            | the clinical periodontal                               |  |  |
| Shfshak e   | CAL.   | parameters evaluated                                   |  |  |
| Ali; 2018   | Parameters evaluated in the FCG:             | significantly compared to the                          |  |  |
|   | Levels of β-defensin-2 (hBD-2).              | control sites, with the                                |  |  |
|   | HA (Gengigel®; 0.8% formulation)             | exception of PI. The                                   |  |  |
|   | was applied subgingivally once just          | expression of hBD-2 was more                           |  |  |
|   | after NSPT.                                  | pronounced in sites that                               |  |  |
|   | Patients were assessed at baseline and       | received subgingival                                   |  |  |
|   | 6 and 12 weeks after NSPT.                   | application of HA in both                              |  |  |
|   |  | reevaluations.   |  |  |
| Lobato et   | Periodontal parameters evaluated:            | HA was able to reduce BoP in                           |  |  |
| al., 2019   | PPD, BoP, PI, GI and CAL.                    | both revaluations. The other                           |  |  |
|   | HA (Gengigel®; 0.8% formulation)             | periodontal parameters, such as                        |  |  |
|   | was applied subgingivally once just          | CAL and PPD, showed a slight                           |  |  |
|   | after NSPT.                                  | improvement compared to the                            |  |  |
|   | Patients were assessed at baseline and       | control sites, but the difference                      |  |  |
|   | 6 and 12 weeks after NSPT.                   | was not statistically significant.                     |  |  |
| Methodology   | : At baseline, patients with periodontities  | s were evaluated for periodontal                       |  |  |
| parameters and were divided into control and test groups, who received, respectively, |  |  |  |  |
| NSPT and placebo or NSPT and subgingival application of HA. Then, patients were       |  |  |  |  |
| reassessed at different times.  |  |  |  |  |
| Eick et al.,  | Periodontal parameters evaluated: PI,        | The HA provided a significant                          |  |  |
| 2013  | BoP, PPD and CAL.                            | reduction in PPD, compared to                          |  |  |
|   |  |  |  |  |
|   | Parameters evaluated in the GCF:             | the control group, but no                              |  |  |
|   | Parameters evaluated in the GCF: MPO levels. | the control group, but no significant differences were |  |  |
|   |  |  |  |  |

|   |  | 1 |  |  |
|---|--|---|--|--|
|   | Campylobacter rectus (Cr),             | in the analysis of MPO. At 6            |  |  |
|   | Aggregatibacter                        | months, it was observed that            |  |  |
|   | actinomycetemcomitans (Aa),            | HA significantly reduced the            |  |  |
|   | Treponema denticola (Td), Prevotella   | microbiota of Aa and Cr in the          |  |  |
|   | intermedia (Pi) and Porphyromona       | test group, compared to the             |  |  |
|   | gingivalis (Pg).                       | control. Despite not having             |  |  |
|   | 0.8% HA (Gengigel®; 0.8%               | interfered with the microbiota          |  |  |
|   | formulation) was applied               | of the other analyzed                   |  |  |
|   | subgingivally once just after NSPT.    | specimens, the HA provided              |  |  |
|   | Then, the 0.2% HA gel (Gengigel®;      | maintenance of its reduced              |  |  |
|   | 0.2% formulation) was applied daily,   | counts after NSPT, in                   |  |  |
|   | twice a day for two weeks, after       | comparison to the control               |  |  |
|   | NSPT.                                  | group.                                  |  |  |
|   | Patients were assessed at baseline, 3  |   |  |  |
|   | and 6 months after NSPT                |   |  |  |
| Sharma et   | Periodontal parameters evaluated: PI,  | HA did not significantly                |  |  |
| al., 2016   | papillary bleeding, gingival staining, | improve, compared to the                |  |  |
|   | PPD and CAL.                           | control group, the parameters           |  |  |
|   | 0.8% HA (Gengigel®; 0.8%               | evaluated, although a trend             |  |  |
|   | formulation) was applied               | towards positive results was            |  |  |
|   | subgingivally once just after NSPT.    | observed.                               |  |  |
|   | Patients were assessed at baseline and |   |  |  |
|   | 1, 2 and 6 weeks after NSPT.           |   |  |  |
| <b>Methodology:</b> In vitro study carried out with human fibroblasts stimulated by Pg. |  |   |  |  |
| Chen et al.,  | Inflammatory responses were induced    | Only high-weight HA (1300               |  |  |
| 2018  | in culture of human gingival           | kDa) reduced the production of          |  |  |
|   | fibroblasts, which were cultured with  | inflammatory mediators by               |  |  |
|   | different molecular weights of HA      | HgFs, induced by Pg,                    |  |  |
|   | (30, 300 and 1300 kDa).                | influencing the inflammation-           |  |  |
|   |  | related NF-κB, ERK and p38              |  |  |
|   |  | MAPK pathways.                          |  |  |

### **4 DISCUSSION**

HA was discovered by Karl Meyer and John Palmer in 1934, after observing a viscous element in the vitreous body of the cow<sup>28</sup>. This name was proposed to him because this substance consists of two types of sugar molecules, D-glucuronic acid (GlcUA) and N-acetylglucosamine (GlcNAc), interconnected by glycosidic bonds. Although it was first identified as an acid, under physiological conditions HA works as sodium hyaluronate<sup>28</sup>, being found mainly in the connective tissue of vertebrates. Especially in humans, it is identified in the umbilical cord,

synovial fluid, skin, vitreous, lungs, kidneys, brain, muscles and, to a lesser extent, in blood plasma<sup>29,30</sup>.

It is a compound produced essentially by fibroblasts in the presence of endotoxins and the main constituent of the extracellular matrix, which is responsible for the growth and function of cells, in addition to their indispensable role in organogenesis. The main physiological function associated with HA involves tissue hydrodynamics and cell migration, proliferation and cohesion<sup>29,30,31,32</sup>.

Such a substance is naturally present in the extracellular space and, therefore, its synthesis occurs on the internal surface of the fibroblast plasma membrane, the result of the action of a protein that binds to the membrane<sup>30,32</sup>. Namely, there are 3 types of HA synthetases found in periodontal tissues that differ in their functions of producing high and medium (HMW and MMW) and low (LMW) molecular weight of HA<sup>31</sup>.

In its soluble form, HA, once in the extracellular matrix, establishes covalent type binding with proteins, which, in turn, express cell receptors, CD44 and RHAMM, both involved in tissue inflammation through the activation of T and lymphocytes. of cell proliferation by mitosis. Interestingly, such receptors are involved in metastases and growth tumors. It is noteworthy that when there is a connection between RHAMM and HA, it has an important role in tissue repair<sup>31,32</sup>. Similarly, the degradation of HA also occurs through the fibroblasts themselves, with the action of the enzyme hyaluronidase during lymphatic drainage into the bloodstream or local metabolism, with the majority of HA being eliminated by the liver and, to a lesser extent, by the kidneys.<sup>30</sup>.

In addition to the hygroscopic and viscoelastic chemical properties, HA has biological characteristics and implications that occur through the interaction of HA with the extracellular matrix<sup>33</sup>. It has been used in situations that demand its healing effect, such as cataract surgery in ophthalmology, dermatology and orofacial aesthetics, being useful in filling the dermis<sup>34</sup>. Namely, the first medical use of HA reported was as a substitute for vitreous during eye surgery. In sequence, for years HA has been shown to be effective as an auxiliary healing agent in the treatment of maxillofacial disorders, in the treatment of temporomandibular disorders, dentin repair process, dental pulp regeneration process, and in periodontics actions, such as in interproximal papilla filling and used as an irrigating solution in subgingival scraping<sup>30,31,33,35</sup>.

In the vast majority of studies in the present review, there were advantages to the administration of HA, under different protocols, in association with SRP<sup>17-25</sup>. In fact, the bone protective and anti-inflammatory effect of HA has been observed in several studies, especially

in those involving osteoarthritis, also considering its lubricating, chondroprotective<sup>12-14,36</sup> and osteoconductive effect<sup>37</sup>. With regard to the latter, the potential of HA for bone regeneration has been highlighted<sup>37</sup>, corroborating the potential healing activity of HA seen in studies that observed an improvement especially of CAL in test sites<sup>20-23</sup>.

After tissue damage occurs in infectious periodontal disease, an induction of inflammation is observed, characterized by the production and release of cytokines, in addition to other chemical substances, such as eicosanoids and growth factors, through inflammatory, fibroblasts and endothelial cells<sup>38</sup>. Endothelial cells, in response to pro-inflammatory cytokines, specifically TNF- $\alpha$ , IL-1 $\beta$  and IL-8, will synthesize HA endogenously in order to counterbalance the inflammatory process, thus contributing to the stabilization of the granulation tissue matrix<sup>38</sup>.

In the granulation phase, HA, in turn, favors tissue repair functions, such as cell migration and proliferation, moderation of the inflammatory response and angiogenesis<sup>39</sup>. The third phase of healing will be the re-epithelialization process. In the latter, the presence of HA in the epidermis has an effect on the elimination of free radicals, in addition to interfering with the migration and proliferation of keratinocytes. Finally, remodeling happens when the granulation tissue evolves into a scar. At this time, HA starts to be reduced in quantity and consequently in effect, giving space for the performance of collagen that offers better increase in the tensile strength of tissues<sup>40</sup>. Therefore, CAL gain after HA administration indicates that HA plays an important role in post-inflammatory tissue regeneration, facilitating cell migration and differentiation during tissue repair.

Despite this, Bevilacqua et al. (2012)<sup>18</sup>, in their research, evaluated the levels of myeloperoxidase (MPO) in the crevicular fluid under the effect of HA and observed that the application of HA did not provide additional benefits in reducing the concentration of this enzyme. In this context, it is worth mentioning that MPO is a marker of polymorphonuclear neutrophil infiltration. Thus, this result does not exclude the anti-inflammatory activity of HA in the spread of the inflammatory process, as it is known that the neutrophil is an expressive cell in acute inflammation, of short duration, whose peak occurs after 48 hours, decreasing soon after<sup>1,2</sup>.

Corroborating these findings, later data reported by Mallikarjun et al.  $(2016)^{21}$  also highlighted that HA did not improve levels of neutrophilic elastase (NE) in the crevicular fluid of patients with PD, compared to control sites. These authors even reported that there was no positive correlation between the improvement of periodontal clinical parameters evaluated after 1 month of NSPT and the levels of NE. Thus, although such results indicate that HA does not

interfere with the positive effect of NSPT on reducing levels of neutrophilic activity, it is suggested that the effect of HA may be more prominent in cells related to the healing process, such as fibroblasts, and that the effect of HA on the expression of matrix metalloproteinases, as well as inflammatory markers associated with macrophage activity, is further explored <sup>18,21</sup>.

In fact, the immunomodulatory effect of HA could be sustained by Chen et al.  $(2018)^{27}$ , in which a reduction in the levels of pro-inflammatory cytokines was observed, via nuclear factor Kappa-B (NF- $\kappa$ B), ERK and (Mitogen-activated protein kinases (MAPK) p38, and human fibroblast cultures induced by *Porphyromonas gingivalis*, which corroborates previous studies in which the use of high molecular weight HA was able to reduce the conversion of macrophages to the M1 phenotype and, consequently, the production of pro-inflammatory mediators. However, although Chen et al.  $(2018)^{27}$  have not observed benefits with low and medium molecular weight HA, it is worth noting that recently it has been observed that low molecular weight HA is also able to influence the release of TNF- $\alpha$ , IL-6, IL-1, as well as pro-inflammatory enzymes, in long-term culture of astrocytes induced by lipopolysaccharide  $^{42}$ .

Finally, some studies in the present literature review analyzed the antimicrobial effect of  $HA^{20,25}$  and obtained promising results. Namely, with regard to its antimicrobial property, the bacteriostatic effect of  $HA^{43}$  has been observed for decades. In fact, more recently, reports have been found on its bacterial and fungal inhibitory effect<sup>30,44,45</sup>. Regarding the formation of the biofilm, it is believed that HA acts to prevent the bacterial adhesion phase, which is necessary for the colonization of the initial colonizers of the biofilm<sup>45</sup>. In addition, it is worth noting that Al-shammari, Shfshak and Ali  $(2018)^{23}$  detected higher levels of  $\beta$ -defensin-2 in sites that received HA administration, compared to control sites. Namely, such defensins are broadspectrum antimicrobial peptides produced by epithelial cells, in addition to important chemotactic functions<sup>46</sup>.

### **5 CONCLUSIONS**

In general, there is a lack of standardization in the protocol for the use of HA in periodontal bags, and the different methodologies that vary both in the amount of gel used and in protocols for different uses, which may change the expected results. It is also recalled that the period between analyzes and monitoring is different in each survey. However, the use of HA as subgingival irrigation as an adjunctive treatment for PD shows promising trends due to its benefits.

#### REFERENCES

Deo V, Bhongade ML. Pathogenesis of periodontitis: role of cytokines in host response. Dent Today. 2010 Sep;29(9):60-2, 64-6; quiz 68-9.

Yucel-Lindberg T, Båge T. Inflammatory mediators in the pathogenesis of periodontitis. Expert Rev Mol Med. 2013 Aug 5;15:e7.

Papapanou PN, Sanz M, Buduneli N, Dietrich T, Feres M, Fine DH, et al. Periodontitis: Consensus report of workgroup 2 of the 2017 World Workshop on the Classification of Periodontal and Peri-Implant Diseases and Conditions. J Periodontol. 2018 Jun;89 Suppl 1:S173-S182.

Loeser RF, Goldring SR, Scanzello CR, Goldring MB. Osteoarthritis: a disease of the joint as an organ. Arthritis Rheum. 2012 Jun;64(6):1697-707.

Aimetti M. Nonsurgical periodontal treatment. Int J Esthet Dent. 2014 Summer;9(2):251-67. Deas DE, Moritz AJ, Sagun RS Jr, Gruwell SF, Powell CA. Scaling and root planing vs. conservative surgery in the treatment of chronic periodontitis. Periodontol 2000. 2016 Jun;71(1):128-39.

Teles RP, Teles FR. Antimicrobial agents used in the control of periodontal biofilms: effective adjuncts to mechanical plaque control? Braz Oral Res. 2009;23 Suppl 1:39-48.

Giurgiu MC, Stana P, Maniu G, Traian DH, Silvia DA. Biotechnological value of the hyaluronic acid in periodontal treatment. Romanian Biotechnological Letters. 2013;18(4):8551-8558.

Sehdev B, Bhongade ML, Ganji KK. Evaluation of effectiveness of hyaluronic acid in combination with bioresorbable membrane (poly lactic acid-poly glycolic acid) for the treatment of infrabony defects in humans: A clinical and radiographic study. J Indian Soc Periodontol. 2016 Jan-Feb;20(1):50-6.

Dalmedico MM, Meier MJ, Felix JV, Pott FS, Petz Fde F, Santos MC. Hyaluronic acid covers in burn treatment: a systematic review. Rev Esc Enferm USP. 2016 May-Jun;50(3):522-8.

Longinotti C. The use of hyaluronic acid based dressings to treat burns: A review. Burns Trauma. 2014 Oct 25;2(4):162-8.

Salamanna F, Giavaresi G, Parrilli A, Martini L, Nicoli Aldini N, Abatangelo G, et al. Effects of intra-articular hyaluronic acid associated to Chitlac (arty-duo®) in a rat knee osteoarthritis model. J Orthop Res. 2019 Apr;37(4):867-876.

Bergstrand S, Ingstad HK, Møystad A, Bjørnland T. Long-term effectiveness of arthrocentesis with and without hyaluronic acid injection for treatment of temporomandibular joint osteoarthritis. J Oral Sci. 2019 Mar 28;61(1):82-88.

Liu SC, Qiao XF, Tang QX, Li XG, Yang JH, Wang TQ, et al. Therapeutic efficacy of extracorporeal shock wave combined with hyaluronic acid on knee osteoarthritis. Medicine (Baltimore). 2019 Feb;98(8):e14589.

Guzińska K, Kaźmierczak D, Dymel M, Pabjańczyk-Wlazło E, Boguń M. Anti-bacterial materials based on hyaluronic acid: Selection of research methodology and analysis of their anti-bacterial properties. Mater Sci Eng C Mater Biol Appl. 2018 Dec 1;93:800-808.

Drago L, Cappelletti L, De Vecchi E, Pignataro L, Torretta S, Mattina R. Antiadhesive and antibiofilm activity of hyaluronic acid against bacteria responsible for respiratory tract infections. APMIS. 2014 Oct;122(10):1013-9.

Pilloni A, Annibali S, Dominici F, Di Paolo C, Papa M, Cassini MA, et al. Evaluation of the efficacy of an hyaluronic acid-based biogel on periodontal clinical parameters. A randomized-controlled clinical pilot study. Ann Stomatol (Roma). 2011 Mar;2(3-4):3-9.

Bevilacqua L, Eriani J, Serroni I, Liani G, Borelli V, Castronovo G, et al. Effectiveness of adjunctive subgingival administration of amino acids and sodium hyaluronate gel on clinical and immunological parameters in the treatment of chronic periodontitis. Ann Stomatol (Roma). 2012 Apr;3(2):75-81.

Gontiya G, Galgali SR. Effect of hyaluronan on periodontitis: A clinical and histological study. J Indian Soc Periodontol. 2012 Apr;16(2):184-92.

Polepalle T, Srinivas M, Swamy N, Aluru S, Chakrapani S, Chowdary BA. Local delivery of hyaluronan 0.8% as an adjunct to scaling and root planing in the treatment of chronic periodontitis: A clinical and microbiological study. J Indian Soc Periodontol. 2015 Jan-Feb;19(1):37-42.

Mallikarjun S, Neelakanti A, Babu HM, Pai SB, Shinde SV, Krishnan S. Neutrophil elastase levels in the gingival crevicular fluid following hyaluronan gel application in the treatment of chronic periodontitis: A randomized split-mouth study. Indian J Dent Res. 2016 Jul-Aug;27(4):397-404.

Shah SA, Vijayakar HN, Rodrigues SV, Mehta CJ, Mitra DK, Shah RA. To compare the effect of the local delivery of hyaluronan as an adjunct to scaling and root planing versus scaling and root planing alone in the treatment of chronic periodontitis. J Indian Soc Periodontol. 2016 Sep-Oct;20(5):549-556.

Al-Shammari NM, Shafshak SM, Ali MS. Effect of 0.8% Hyaluronic Acid in Conventional Treatment of Moderate to Severe Chronic Periodontitis. J Contemp Dent Pract. 2018 May 1;19(5):527-534.

Lobato JCRF, Dos Santos Vilhena MA, Izidoro C, Alves RC, Proença L. Single application of 0.8% **hyaluronic acid** as a coadjuvant of nonsurgical treatment in nonsmoking patients with **periodontitis**: A split-mouth, randomized, controlled pilot clinical trial. J Indian Soc Periodontol. 23(6):545-548. 2019.

Eick S, Renatus A, Heinicke M, Pfister W, Stratul SI, Jentsch H. Hyaluronic Acid as an adjunct after scaling and root planing: a prospective randomized clinical trial. J Periodontol. 2013 Jul;84(7):941-9.

Sharma V, Gupta R, Dahiya P, Kumar M. Comparative evaluation of coenzyme Q10-based gel and 0.8% hyaluronic acid gel in treatment of chronic periodontitis. J Indian Soc Periodontol. 2016 Jul-Aug;20(4):374-380.

Chen M, Li L, Wang Z, Li P, Feng F, Zheng X. High molecular weight hyaluronic acid regulates P. gingivalis-induced inflammation and migration in human gingival fibroblasts via MAPK and NF-κB signaling pathway. Arch Oral Biol. 2019 Feb;98:75-80.

Meyer K, Palmer JW. The Polysaccharide of the Vitreous Humor. J. Biol. Chem. 1934;107(1):629–34.

Liu L, Liu Y, Li J, Du G, Chen J. Microbial production of hyaluronic acid: current state, challenges, and perspectives. Microb Cell Fact. 2011 Nov 16;10:99. Dahiya P, Kamal R. Hyaluronic Acid: a boon in periodontal therapy. N Am J Med Sci. 2013

May;5(5):309-15.

Jiang D, Liang J, Noble PW. Hyaluronan as an immune regulator in human diseases. Physiol Rev. 2011 Jan;91(1):221-64.

Shimabukuro Y, Terashima H, Takedachi M, Maeda K, Nakamura T, Sawada K, et al. Fibroblast growth factor-2 stimulates directed migration of periodontal ligament cells via PI3K/AKT signaling and CD44/hyaluronan interaction. J Cell Physiol. 2011 Mar;226(3):809-21.

Bansal J, Kedige SD, Anand S. Hyaluronic acid: a promising mediator for periodontal regeneration. Indian J Dent Res. 2010 Oct-Dec;21(4):575-8.

Salwowska NM, Bebenek KA, Żądło DA, Wcisło-Dziadecka DL. Physiochemical properties and application of hyaluronic acid: a systematic review. J Cosmet Dermatol. 2016 Dec;15(4):520-526.

Cömert Kiliç S, Güngörmüş M. Is arthrocentesis plus platelet-rich plasma superior to arthrocentesis plus hyaluronic acid for the treatment of temporomandibular joint osteoarthritis: a randomized clinical trial. Int J Oral Maxillofac Surg. 2016 Dec;45(12):1538-1544.

Bowman S, Awad ME, Hamrick MW, Hunter M, Fulzele S. Recent advances in hyaluronic acid based therapy for osteoarthritis. Clin Transl Med. 2018 Feb 16;7(1):6.

Yeom J, Hwang BW, Yang DJ, Shin HI, Hahn SK. Effect of osteoconductive hyaluronate hydrogels on calvarial bone regeneration. Biomater Res. 2014 Jul 23;18:8.

**Gurtner** GC, Werner S, Barrandon Y, Longaker MT. **Wound repair and regeneration**. Nature. 2008 May 15;453(7193):314-21.

Dicker KT, Gurski LA, Pradhan-Bhatt S, Witt RL, Farach-Carson MC, Jia X. Hyaluronan: a simple polysaccharide with diverse biological functions. Acta Biomater. 2014 Apr;10(4):1558-70.

Reinke JM, Sorg H. Wound repair and regeneration. Eur Surg Res. 2012;49(1):35-43

Bauer C, Niculescu-Morzsa E, Jeyakumar V, Kern D, Späth SS, Nehrer S. Chondroprotective effect of high-molecular-weight hyaluronic acid on osteoarthritic chondrocytes in a co-cultivation inflammation model with M1 macrophages. J Inflamm (Lond). 2016 Sep 13;13(1):31.

Chistyakov DV, Astakhova AA, Azbukina NV, Goriainov SV, Chistyakov VV, Sergeeva MG. **High** and **Low Molecular Weight Hyaluronic Acid Differentially Influences Oxylipins Synthesis** in **Course** of **Neuroinflammation**. Int J Mol Sci. 2019 Aug 9;20(16). pii: E3894.

Pirnazar P, Wolinsky L, Nachnani S, Haake S, Pilloni A, Bernard GW. Bacteriostatic effects of hyaluronic acid. J Periodontol. 1999 Apr;70(4):370-4.

Rodrigues SV, Acharya AB, Bhadbhade S, Thakur SL. Hyaluronan-containing mouthwash as an adjunctive plaque-control agent. Oral Health Prev Dent. 2010;8(4):389-94.

Romanò CL, De Vecchi E, Bortolin M, Morelli I, Drago L.**Hyaluronic Acid** and Its Composites as a Local Antimicrobial/Antiadhesive Barrier. J Bone Jt Infect. 2017 Jan 1;2(1):63-72.

Niyonsaba F, Ogawa H, Nagaoka I. Human beta-defensin-2 functions as a chemotactic agent for tumour necrosis factor-alpha-treated human neutrophils. Immunology. 2004 Mar;111(3):273-81.